Amendments to the Specification:

Please replace the paragraph at page 11, lines 22-23, with the following amended paragraph:

Figure 3C. Amino Acid Sequences of the Constant Regions of the Modified OST577-IgG1 in Comparison with the Unmodified OST577-IgG1 (SEQ ID NO: 3). The amino acid sequences of the constant region of the heavy chain of the modified IgG1 having the amino acid substitution of T250D (SEQ ID NO: 67), T250E (SEQ ID NO: 68), T250Q (SEQ ID NO: 69), M428F (SEQ ID NO: 70), M428L (SEQ ID NO. 71), T250E/M428F (SEQ ID NO: 75), and T250Q/M428L (SEQ ID NO: 76) are also shown.

Please replace the paragraph at page 12, lines 3-4, with the following amended paragraph:

Figure 3E. Amino Acid Sequences of the Constant Regions of the Modified Hu1D10-IgG2M3 in Comparison with the Unmodified Hu1D10-IgG2M3. The IgG2M3 sequence is represented by SEQ ID NO:2. M428L is represented by SEQ ID NO:57; T250Q/M428L is represented by SEQ ID NO: 74.

Please replace the paragraph at page 12, lines 5-6, with the following amended paragraph:

Figure 3F. Amino Acid Sequences of the Constant Regions of the Modified Hu1D10-IgG1 in Comparison with the Unmodified Hu1D10-IgG1. The IgG1 sequence is represented by SEQ ID NO:3. M428L is represented by SEQ ID NO:71; T250Q/M428L is represented by SEQ ID NO: 76.

Please replace the paragraph at page 12, lines 16-17, with the following amended paragraph:

Figure 3H. Amino Acid Sequences of the Constant Regions of the M428L Hu1D10-IgG3 Mutant (SEQ ID NO: 115) in Comparison with the Unmodified Hu1D10-IgG3 (SEQ ID NO:113).

Please replace the paragraph at page 12, lines 18-19, with the following amended paragraph:

Figure 3I. Amino Acid Sequences of the Constant Regions of the M428L Hu1D10-IgG4 (SEQ ID NO:116) and T250Q/M428L Hu1D10-IgG4 (SEQ ID NO:117) Mutants in Comparison with the Unmodified Hu1D10-IgG4 (SEQ ID NO:114).

Please replace the paragraph at page 15, lines 15-17, with the following amended paragraph:

Figure 17A. Binding Assay of Hu1D10-IgG2M3 Wild-Type and Mutant Antibodies to HLA-DR β Chain Allele. The binding of the wild-type or mutant Hu1D10-IgG2M3 antibodies to Raji cells was analyzed in a FACS™ binding assay, as described in Example 8.

Please replace the paragraph at page 15, lines 18-20, with the following amended paragraph:

Figure 17B. Binding Assay of Hu1D10-IgG1 Wild-Type and Mutant Antibodies to HLA-DR β Chain Allele. The binding of the wild-type or mutant Hu1D10-IgG1 antibodies to Raji cells was analyzed in a FACS™ binding assay, as described in Example 8.

Please replace the paragraph at page 65, lines 4-24, with the following amended paragraph:

NS0 cells were stably transfected with pDL208. Approximately $1x10^7$ cells were washed once and resuspended in 1 ml of plain DMEM, transferred to a Gene Pulser™ Cuvette (Bio-Rad® Laboratories), and incubated on ice for 10 minutes. Forty μg of plasmid pDL208 was

linearized with FspI and gently mixed with the cells on ice, then the cells were electroporated by pulsing twice using a Gene Pulser™ II (Bio-Rad® Laboratories) set at 1.5 kV, 3 μF, and returned to ice for 10 minutes. The cells were diluted in 20 ml of DMEM, 10% FBS, and plated in two 96-well plates at 100 μl/well. The medium was replaced after 48 hours with MPA selection medium. Mycophenolic acid-resistant NS0 transfectants from wells apparently containing single colonies were expanded in MPA selection medium and screened after about 3 weeks by FACS™. Approximately $1.5x10^5$ cells/test were incubated in 100 μl of FACS Staining Buffer (FSB) (PBS, 1% FBS, 0.1% NaN_3) containing 10 μg/ml of biotinylated mouse anti-human β2-microglobulin antibody (Chromaprobe, Inc., Aptos, Calif.) for 1 hour on ice. The cells were washed once with 4 ml of FSB, then incubated in 25 μl of FSB containing 20 μg/ml of streptavidin-FITC conjugate (Southern Biotechnology Associates, Inc.) for 30 minutes on ice in the dark. The cells were washed once with 4 ml of FSB, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to human β2m using a FACScan™ flow cytometer (BD® Biosciences, San Jose, Calif.). Several clones with the highest apparent staining were subcloned using a FACStar cell sorter (BD® Biosciences), expanded in DMEM, 10% FBS, 2 mM L-glutamine, and retested by FACS™ as described above. One subclone, designated NS0 HuFcRn (memb), clone 7-3, was used in subsequent binding assays.

Please replace the paragraph at page 66, lines 2-14, with the following amended paragraph:

Concentrated OST577-IgG2M3 supernatants were tested in a single-point competitive binding assay for binding to human FcRn on cell line NS0 HuFcRn (memb), clone 7-3. Approximately $2x10^5$ cells/test were washed once in FACS Binding Buffer (FBB) (PBS containing 0.5% BSA, 0.1% NaN_3), pH 8.0, once in FBB, pH 6.0, and resuspended in 120 μl of pre-mixed biotinylated OST577-IgG2M3 antibody (8.3 μg/ml) and concentrated supernatant (containing 8.3 μg/ml of competitor antibody) in FBB, pH 6.0. he cells were incubated for 1 hour on ice, washed twice in FBB, pH 6.0, and resuspended in 25 μl of streptavidin-RPE conjugate (BioSource International) diluted to 2.5 μg/ml in FBB, pH 6.0. After incubation for 30 minutes

on ice in the dark, the cells were washed twice in FBB, pH 6.0, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACSCalibur™ flow cytometer (BD® Biosciences). Mean channel fluorescence (MCF) of each mutant was compared to that of the wild-type antibody and plotted using Excel (Microsoft® Corporation, Redmond, Wash.).

Please replace the paragraph at page 66, lines 16-30, with the following amended paragraph:

A dilution series of each purified OST577-IgG2M3 antibody was competed against biotinylated HuEP5C7-IgG2M3 antibody (He et al., J. Immunol. 160:1029-1035 (1998)) for binding to human FcRn on cell line NS0 HuFcRn (memb), clone 7-3. For initial screening experiments, approximately 2×10^5 cells/test were washed once in FSB, pH 6.0, and resuspended in 100 µl of pre-mixed biotinylated HuEP5C7-IgG2M3 antibody (10 µg/ml) and OST577-IgG2M3 competitor antibody (twofold serial dilutions from 208 µg/ml to 0.102 µg/ml) in FSB, pH 6.0. The cells were incubated with the antibody mixture for 1 hour on ice, washed twice in FSB, pH 6.0, and resuspended in 25 µl of streptavidin-RPE conjugate (BioSource International) diluted to 2.5 µg/ml in FSB, pH 6.0. After incubation for 30 minutes on ice in the dark, the cells were washed twice in FSB, pH 6.0, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACScan™ flow cytometer (BD® Biosciences). Mean channel fluorescence (MCF) was plotted against competitor concentration, and IC50 values were calculated using GraphPad Prism® (GraphPad™ Software, Inc., San Diego, Calif.). For consistency, the IC50 values shown in the Tables are based on the final competitor concentrations.

Please replace the paragraph at page 67, lines 12-26, with the following amended paragraph:

A dilution series of each purified OST577-IgG2M3 antibody was competed against biotinylated OST577-IgG2M3 antibody for binding to rhesus FcRn on cell line NS0 RhFcRn, clone R-3. In one group of experiments, approximately 2×10^5 cells/test were washed once in FBB, pH 8.0, and once in FBB, pH 6.0, then resuspended in 120 µl of pre-mixed biotinylated OST577-IgG2M3 antibody (8.3 µg/ml) and OST577-IgG2M3 competitor antibody (twofold serial dilutions from 208 µg/ml to 0.102 µg/ml) in FBB, pH 6.0. The cells were incubated with the antibody mixture for 1 hour on ice, washed twice in FBB, pH 6.0, and resuspended in 25 µl of streptavidin-RPE conjugate (BioSource International) diluted to 2.5 µg/ml in FBB, pH 6.0. After incubation for 30 minutes on ice in the dark, the cells were washed twice in FBB, pH 6.0, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACSCalibur™ flow cytometer (BD® Biosciences). Another group of experiments was done in 200 µl of pre-mixed biotinylated OST577-IgG1 antibody (5.0 µg/ml) and OST577-IgG1 competitor antibody (threefold serial dilutions starting from 500 µg/ml) in FBB, pH 6.0, as described above.

Please replace the paragraph beginning at page 73, lines 4-14, with the following amended paragraph:

Purified OST577-IgG2M3 antibodies were tested for binding to human FcRn on cell line NS0 HuFcRn (memb), clone 7-3, or to untransfected NS0 cells in FBB at pH 6.0. Approximately 2×10^5 cells/test were washed once in FBB, pH 8.0, and once in FBB, pH 6.0, then resuspended in 100 µl of antibody at a concentration of 11 µg/ml in FBB, pH 6.0. The cells were incubated with antibody for 1 hour on ice, washed twice in FBB, pH 6.0, and resuspended in 25 µl of goat anti-human IgG RPE-conjugated antibody (Southern Biotechnology Associates, Inc.) diluted to 5 µg/ml in FBB, pH 6.0. After incubation for 30 minutes on ice in the dark, the cells were washed twice in FBB, pH 6.0, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACScan™ flow cytometer (BD®

Biosciences). Mean channel fluorescence (MCF) of each mutant was plotted using Excel (Microsoft® Corporation).

Please replace the paragraph at page 73, lines 14-29, with the following amended paragraph:

A dilution series of each purified OST577-IgG2M3 antibody was competed against biotinylated OST577-IgG2M3 antibody for binding to human FcRn on cell line NS0 HuFcRn (memb), clone 7-3 at 37° C. Approximately 2×10^5 cells/test were washed once in FBB, pH 8.0, and once in FBB, pH 6.0, then resuspended in 100 µl of pre-mixed biotinylated OST577-IgG2M3 antibody (10 µg/ml) and OST577-IgG2M3 competitor antibody (twofold serial dilutions, from 208 µg/ml to 0.102 µg/ml) in FBB, pH 6.0. The cells were incubated with the antibody mixture for 1 hour at 37° C., washed twice in FBB, pH 6.0, and resuspended in 25 µl of streptavidin-RPE conjugate (BioSource International) diluted to 2.5 µg/ml in FBB, pH 6.0. After incubation for 30 minutes in the dark, the cells were washed twice in FBB, pH 6.0, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACScan™ flow cytometer (BD® Biosciences). Mean channel fluorescence (MCF) was plotted against competitor concentration, and IC50 values were calculated using GraphPad Prism® (GraphPad™ Software).

Please replace the paragraph beginning at page 73, line 31, with the following amended paragraph:

Purified OST577-IgG2M3 and OST577-IgG1 mutant antibodies were compared to the respective wild-type antibodies for binding to human FcRn and then released at various pH values in single-point binding and release assays using cell line NS0 HuFcRn (memb), clone 7-3. Approximately 2×10^5 cells/test were washed once in FBB, pH 8.0, and once in FBB, pH 6.0, then resuspended in 100 µl of purified antibody (10 µg/ml) in FBB, pH 6.0. The cells were

incubated for 1 hour on ice, washed twice in FBB, pH 6.0, 6.5, 7.0, 7.5, or 8.0, and resuspended in 25 μl of goat $F(ab')_2$ anti-human IgG FITC-conjugated antibody (Southern Biotechnology Associates, Inc.) diluted to 1.25 μg/ml in FBB of the appropriate pH. After incubation for 30 minutes on ice in the dark, the cells were washed twice in FBB of the appropriate pH, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACSCalibur™ flow cytometer (BD® Biosciences). Mean channel fluorescence (MCF) of each mutant was plotted using Excel (Microsoft® Corporation).

Please replace the paragraph beginning at page 76, line 28, with the following amended paragraph:

The antigen binding activity of Hu1D10-IgG2M3 wild-type and mutant antibodies was confirmed in a FACS™ binding assay using Raji cells, which express an allele of the HLA-DR β chain that is recognized by Hu1D10 (Kostelny et al. (2001), op. cit.). Approximately 2.5×10^5 cells/test were washed once in FBB, pH 7.4, and resuspended in 140 μl of Hu1D10-IgG2M3 antibody (threefold serial dilutions from 60 μg/ml to 0.027 μg/ml) in FBB, pH 7.4. The cells were incubated with antibody for 1 hour on ice, washed twice in FBB, pH 7.4, and resuspended in 25 μl of goat $F(ab')_2$ anti-human kappa RPE-conjugated antibody (Southern Biotechnology Associates, Inc.) diluted to 10 μg/ml in FBB, pH 7.4. After incubation for 30 minutes on ice in the dark, the cells were washed twice in FBB, pH 7.4, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to the HLA-DR β chain allele by FACS™ using a FACSCalibur™ flow cytometer (BD® Biosciences).

Please replace the paragraph at page 77, lines 7-19, with the following amended paragraph:

Similarly, the antigen binding activity of Hu1D10-IgG1 wild-type and mutant antibodies was confirmed in a FACS™ binding assay using Raji cells. Approximately 2.0×10^5 cells/test

were washed once in FBB, pH 7.4, and resuspended in 100 µl of Hu1D10-IgG1 antibody (twofold serial dilutions from 25 µg/ml to 12.5 µg/ml, then threefold serial dilutions from 12.5 µg/ml to 0.0020 µg/ml) in FBB, pH 7.4. A dilution series of HuFd79-IgG1 antibody (Co et al., Proc. Natl. Acad. Sci. 88:2869-2873 (1991)) was prepared as described above and used as a negative control. The cells were incubated with antibody for 1 hour on ice, washed twice in FBB, pH 7.4, and resuspended in 25 µl of goat $F(ab')_2$ anti-human IgG FITC-conjugated antibody (Southern Biotechnology Associates, Inc.) diluted to 20 µg/ml in FBB, pH 7.4. After incubation for 30 minutes on ice in the dark, the cells were washed twice in FBB, pH 7.4, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to the HLA-DR β chain allele by FACS™ using a FACSCalibur™ flow cytometer (BD® Biosciences).